

The effect of black currant selenium nanoparticles on dyslipidemia and oxidant-antioxidant status in D- galactose treated rats

Masar Jabbar Jary Al-Kurdy¹ Khalisa Khadim khudair²

1Department of Nursing Techniques, Technical Institute in Al-Diwaniyah, AL-Furat Al-Awsat Technical University, Iraq.

2 Department of Physiology, biochemistry and Pharmacology, College of Veterinary Medicine, University of Baghdad, Iraq.

Corresponding author: E- mail: rosmaso@yahoo.com, Department of Nursing Techniques, Technical Institute in Al-Diwaniyah, AL-Furat Al-Awsat Technical

Received date: 2May2020 Accepted: (466) 8June2020 page: (23-38) Published: 30June2020

Abstract:

The current study was aimed to explore the effect of black currant selenium nanoparticles (BCSeNPs) on serum lipid profile and oxidant- antioxidant state markers in blood of D-galactose (D-gal) treated rats. The green synthesis of SeNPs as described earlier in our previous study was characterized by color changes; Ultraviolet- visible (UV-VIS) spectroscopy; scanning electron microscopy (SEM) techniques; X-ray diffraction analysis (XRD); Fourier transform infrared spectroscopy (FTIR). The results revealed prevalence of brick- red color of BCSeNPs characterized by spherical crystals with average particle size in the range of 18-50 nm. Thirty-two (32) adult male rats were divided randomly and equally into four experimental groups (8/group) and handles as follows for eight weeks: Control (C) group: rats in this group were treated with intra peritoneal injection (*i.p*) and oral intubation of normal saline. T1 group: animals in this group were subjected to *i.p*. of D gal a dose (150 mg/kg/day), which was dissolved in normal saline solution. T2 group: the rats were oral intubation BCSeNPs (1mg/Kg. B. W). T3 group: rats in this group were administered BCSeNPs concurrently with *i.p*. of D-gal in the same previous doses. Blood samples were collected from heart by cardio puncture technique at 2nd and 8th weeks of the experiment and serum samples were used for estimation of some biochemical parameters related to oxidant-antioxidant status markers: Malondialdehyde (MDA) and Total antioxidant capacity (TAO-C); Serum lipid profile: concentration of total cholesterol (TC), triglyceride (TAG), high density lipoprotein- cholesterol (HDL-c), low density lipoprotein- cholesterol (LDL-c), and very low density lipoprotein- cholesterol (VLDL-c). At the end of experiment, after animal scarifying, section from liver was taken for detection of gene expression of glutathione peroxidase.

The results of here in study confirmed a case the oxidative stress and hyperlipidemia in T1 group manifested by significant depression in serum TAO-C concentration and decrease in glutathione peroxidase gene expression level. While, caused elevation in serum MDA concentration and significant decrease in serum concentration of HDL-c with significant elevation in serum concentration of TAG, VLDL-C, TC, and LDL-c. At the end of experiment, BCSeNPs intubation in T2 group caused alleviation of previous mentioned parameters related to oxidative stress and lipid profile. **On conclusion**, the result in a current study showed that black currant selenium nanoparticle has both a preventive and a therapeutic role in D-gal toxicity in adult male rats, where BCSeNPs can be considered as antioxidant and hypolipidemic agent.

Key Words: Selenium nanoparticles, D-galactose, lipid profile, glutathione peroxidase.

تأثير جسيمات السيلينيوم النانوية- الزبيب الأسود على حالة اضطراب الدهون و حالة الاكسدة - مضادة الاكسدة في لجرذان المعالجة بـ D-gal كالاكتوز

مسار جبار جري الكردي¹ خالصة كاظم خضير²

¹ قسم تقنيات التمريض ، المعهد التقني في الديوانية ، جامعة الفرات الاوسط التقنية ، العراق
² فرع الفلسفة والكيمياء الحياتية والادوية ، كلية الطب البيطري ، جامعة بغداد ، العراق.

المؤلف المقابل: البريد الإلكتروني: rosmaso@yahoo.com ، قسم تقنيات التمريض ، المعهد التقني في الديوانية ، جامعة الفرات الأوسط الفنية

الخلاصة:

هدفت الدراسة الحالية لمعرفة تأثير جسيمات السيلينيوم النانوية المصنعة باستخدام الزبيب الأسود (BCSeNPs) على الصورة الدموية للدهون وبعض المعايير الخاصة بالأكسدة ومضادة الأكسدة في دم جرذان المعالجة بـ D-gal. تم تمييز التصنيع الحيوي لجسيمات السيلينيوم النانوية المرتبطة مع الزبيب الأسود بواسطة تغيرات اللون؛ استخدام التحليل الطيفي المرئي للأشعة فوق البنفسجية (UV-VIS)؛ تقنيات المسح المجهر الإلكتروني (SEM)؛ تحليل حيود الأشعة السينية (XRD)؛ التحليل الطيفي بالأشعة تحت الحمراء فوريريه (FTIR). أوضحت النتائج ظهور لون أحمر قرميد من BCSeNPs يتميز بالبلورات الكروية بمتوسط حجم الجسيمات في حدود 18-50 نانومتر. تم تقسيم اثنان وثلاثون (32) من ذكور الجرذان ويستير البيضاء البالغة بوزن (200 ± 10 غم) بشكل عشوائي وبالتساوي الى أربع مجموعات (ثمانية/مجموعة) وكان تعامل يومياً ولمدة ثمانية اسابيع كالتالي: مجموعة السيطرة (C) عولجت الجرذان في هذه المجموعة عن طريق الحقن داخل الصفاق والتجريب الفموي بمحلول ملح طبيعي. مجموعة T1: تعرضت الجرذان في هذه المجموعة للحقن داخل الصفاق بجرعة (150 ملغ/كغم من وزن الجسم) والذي تم اذابته في محلول ملحي طبيعي. في حين تم تجريب الجرذان في مجموعة T2 جسيمات السيلينيوم النانوية فمويًا بجرعة (1 ملغ/كغم من وزن الجسم). اما حيوانات المجموعة T3 فقد جرعت فمويًا جسيمات السيلينيوم النانوية وحقن د-كالاكتوز داخل الصفاق بنفس الجرعة المذكورة سابقاً. تم جمع عينات الدم عن طريق تقنية ثقب القلب في الأسبوعين الثاني والثامن من التجربة ، وتم تحضير عينات المصل لقياس الاختبارات البيوكيميائية المتعلقة بمؤشرات حالة الأكسدة والمضادة للأكسدة: المالوندايالدهايد (MDA) والسعة الكلية لمضادة الأكسدة (TAO-C)؛ الصورة الدموية للدهون: تركيز الكوليسترول الكلي (TC) ، الدهون الثلاثية (TAG) ، الكوليسترول في البروتين الدهني عالي الكثافة (HDL-C) ، الكوليسترول في البروتين الدهني منخفض الكثافة (LDL-C) ، الكوليسترول في البروتين الدهني الواطئ الكثافة جدا (VLDL-C). تم التضحية بالحيوانات في نهاية التجربة ، تم أخذ جزء من الكبد للكشف عن التعبير الجيني للكلوتاثيون بيروكسيداز. أكدت النتائج هنا في الدراسة حالة الإجهاد التأكسدي وفقرت شحميات الدم في مجموعة T1 يتجلى في انخفاض كبير في تركيز TAO-C في المصل وانخفاض مستوى التعبير الجيني للكلوتاثيون بيروكسيداز. بينما تسبب ارتفاع في تركيز MDA في المصل وانخفاض كبير في تركيز HDL-C مع ارتفاع كبير في تركيز المصل لـ TAG و VLDL-C و TC و LDL-C في نهاية التجربة ، في حين تسبب تجريب BCSeNPs في مجموعة T2 في تقليل حدة المعايير السابقة الذكر المتعلقة بالإجهاد التأكسدي وصورة الدهون. نستنتج من الدراسة الحالية الدور الوقائي والعلاجي لجسيمات السيلينيوم النانوية-الزبيب الأسود ضد سمية D-gal في ذكور الجرذان البالغة، حيث يمكن اعتبار BCSeNPs كعامل مضاد للأكسدة وخافض للدهون .

الكلمات المفتاحية: جسيمات السيلينيوم النانوية ، د-كالاكتوز ، الصورة الدهنية ، الكلوتاثيون بيروكسيداز

Introduction

Nanotechnology sciences provide the improvement of experimental practice for the preparation of the nanoscale constituents with exceptional possessions (1). Today, production of nanoparticles (NPs) using biosynthetic techniques, has been considered as a valuable method with increasing attraction (2,3). Biogenic synthesis of Se nanoparticles is frequently achieved by reduction of selenate/selenite in presence of bacterial proteins or plant extracts containing phenols, flavonoids amines, alcohols, proteins and aldehydes (4,5). Nano-Se possesses better antioxidant capability than other chemical forms of selenium while reducing the risk of selenium toxicity. They appear to have

physiological properties, of important antioxidant effect (6,7). Compound SeNPs protected the liver and kidney against acetaminophen toxicity through reducing oxidative stress, enhancing endogenous antioxidants and protecting mitochondrial functions (8). Its role in treatment of liver disease and as antidiabetic is well documented (9,10). Treatment by SeNPs, is essential to improve health and performance, oral SeNPs supplements showed no disadvantages and its well tolerated by all patients (11). The role of selenium nanoparticles in mitigation of high temperature- stress is enhanced by their antioxidant defense system (12). D-gal is a reducing sugar and can be metabolized at

normal concentration. However, at high levels, it induces the production of reactive oxygen species (ROS) and advanced glycation end products (AGEs) (13,14). It has been suggested that AGEs binding to its receptor form advanced glycation end products (RAGE) in many cell types induces pathophysiological cascades linked to the downstream activation of NF- κ B and other signaling pathways that lead to ROS generation and certain proinflammatory responses (15,16,17). Besides, ROS generation by D-gal could induced memory, systemic dysfunction and neuroinflammation (18). The role of D-gal in inducing aging associated change such as increased oxidative stress, decreased antioxidant enzyme activity and mitochondrial function (19-23) has been well illustrated.

2- Materials and Methods:

Green synthesis of BCSeNPs: using black currant aqueous extract was prepared as described by (24,25). Characterization of BCSeNPs were performed by: Ultraviolet-visible spectroscopy (Metertech SP-8001 Taiwan) as described by (26,27); X-ray diffraction (Shemadzu-6000 Japan) as describe by (28,29); Scanning Electron Microscope (SEM-Tescan Vega III, Czech) as described by (30); Fourier-transform infrared spectroscopy (Shimadzu8400s, Japan) as described by (31,32). The current study had executed in the animal house vassal to the college of Veterinary Medicine, AL-Qadisiya University through the period expanded from January, 2019 to march, 2019. Mature male Wistar rats (aged: 90 days, weighted: 190 ± 5.5 g) have been utilized in the current study. After acclimatization, thirty two (32) adult male rats were divided randomly and equally into four experimental groups and handles as follows for eight weeks: control(C) group : rats in this group were treated with intra peritoneal injection and oral intubated of normal saline, D-galactose(D-gal) (T1) group: animals in this group were subjected to intra peritoneal injection of D-gal a dose (150 mg/kg/day), black currant-selenium nanoparticles(BCSeNPs)(T2)group: the rats were intubation black currant-selenium

nanoparticles (1mg/Kg.B.W) and (T3)group: rats in this group were administered BCSeNPs concurrently with D-gal in the same previous methods and doses. Blood samples were collected by cardiac puncture technique from anesthetized rats, then serum were obtained for measuring the following: Lipid profile including Serum concentration of Total Cholesterol using TC kit (Biosystem Spain), according to (33); Triglyceride utilizing Triglyceride kit (Biosystem .Spain), according to (34); low density lipoprotein-cholesterol and Very-low density lipoprotein-cholesterol depending on Friedewald formula(35) and High density lipoprotein-cholesterol by utilizing HDL-c kit (Biosystem . Spain), according to (36). Besides, blood sample were also obtained for measuring Total antioxidant capacity Kit (Elabscience, USA) and serum MDA by using Thiobarbituric acid (TBA) according to (37). Parts of the liver tissues were removed to detection of Gene expression of glutathione peroxidase (GSH-Px) using RNazol® (Bioneer, korea) was used to extract total RNA from liver tissue, forward primer(5'AGT TCG GAC ATC AGG AGA ATG GCA'3) and Reverse (5'TCA CCA TTC ACC TCG CAC TTC TCA'3) primer used in quantification of gene expression using qRT-PCR techniques based SYBER Green DNA binding dye, and supported from (Bioneer, Korea) company, RT-PCR were identified according to criteria described by (38).Statistical analysis: two-way analysis of various ANOVA and Least significant differences (LSD) test utilizing as prorated (39) at level of ($P < 0.05$).

3-Results:

Green synthesis and characterization of Black currant selenium nanoparticles revealed appearance of brownish after 30 minute that changed gradually to reddish color and it becomes more stable (Figure 1). The optical absorbance of synthesized BcSeNPs was measured using UV-Vis spectroscopy. An absorption peak between (265-370 nm) confirms the presence of BCSeNPs (Figure 2). The pattern of SEM showed spherical shape nanoparticles with a diameter range of

18-50 nm in electron microscope (Figure 3). According to the result of XRD analysis in a current study the physical characteristic of particles in prepared compound is spherical and crystalized nanoparticles and the size of crystal was in range of 18 to 50 nm (Figure 4). Different distinct peak observed in Figure (5) in FTIR analysis, indicated the different functional group present in BcSeNPs. The distinct peak of BcSeNPs was seen at 3352.39 cm^{-1} correspond to OH: NH due to stretch vibration in amide A. Absorption peak at 2931.90 cm^{-1} correspond to C-H in $-\text{CH}_2$ in

aliphatic compounds. While, the band at 1608.69 cm^{-1} indicating NH_2 in primary amides. The peak at 1514.17 cm^{-1} is due to NH in secondary amides (amide II). The peak at 1359.86 cm^{-1} attributed to the C-H bending in alkanes. While, 1066.67 and 1035.81 cm^{-1} confirm C-O, C-C stretching vibrations, C-O-H, C-O-C bending vibrations in polysaccharides, protein and polyesters. C-X stretching in alkyl halides causes a band at 871.85 and 835.21 cm^{-1} . The band at 590.24 and 547.80 cm^{-1} is the result of C-N-C bending in amines

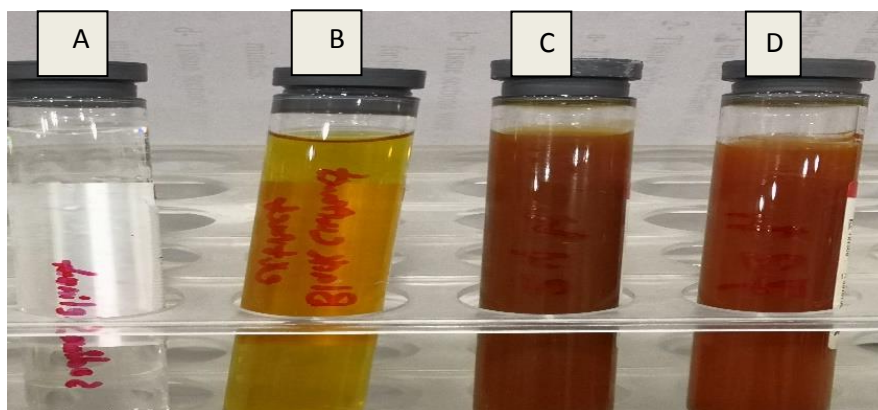


Figure 1: Image showed changing in color after the reduction of sodium selenite to BcSeNPs by black currant extract. **A:** Image showed sodium selenite solution, **B:** Image showed Black currant aqueous extract, **C:** Image showed Black currant selenium nanoparticles, **D:** Image showed BcSeNPs after 48-72 hour.

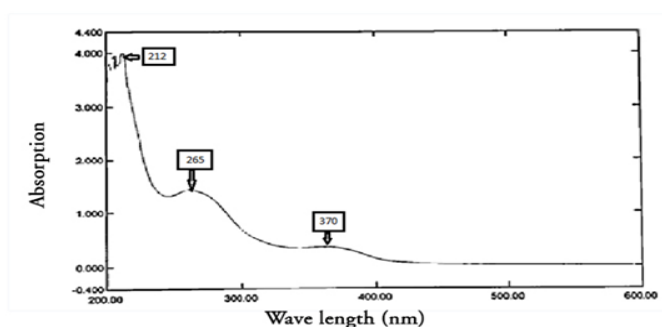


Figure 2. UV-Vis spectroscopy absorbance of selenium nanoparticles, making under carrying out sodium selenite with black currant extract in PH 9.

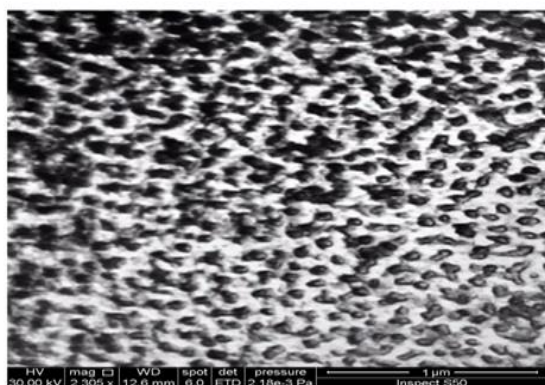


Figure 3. SEM test image of the selenium nanoparticles, making under carrying out sodium selenite with black currant extract in percentage 1:2 v: v ratio in PH 9 (1 μ m)

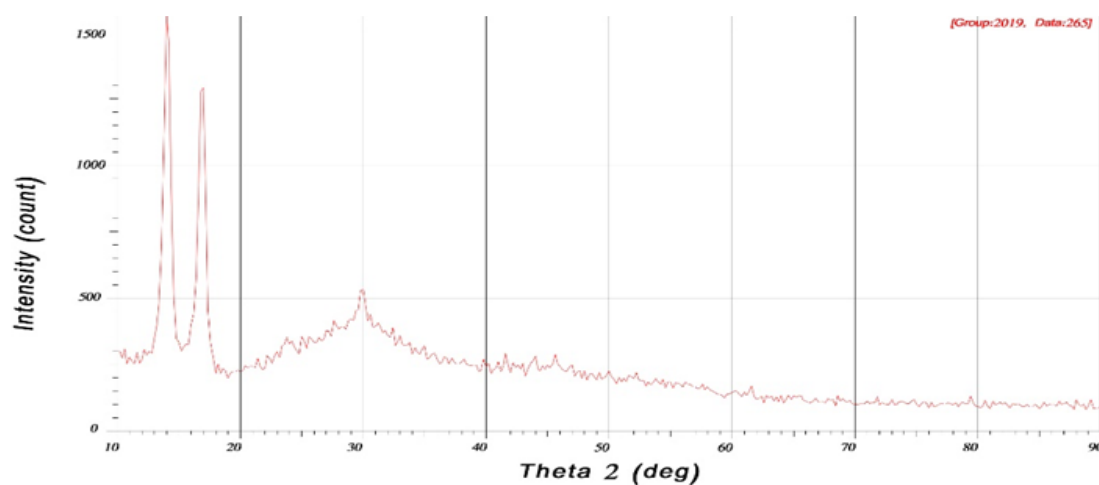


Figure 4: X-ray diffraction pattern for selenium nanoparticles, making under carrying out sodium selenite with black currant extract in PH 9.

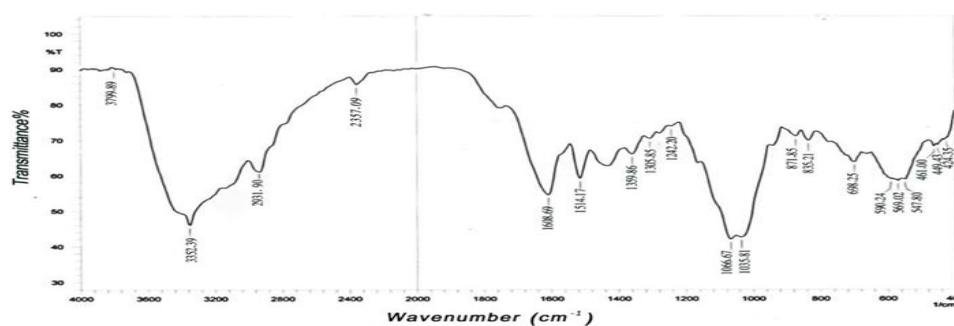


Figure 5: F-TIR spectroscopy for selenium nanoparticles, making under carrying out sodium selenite with black currant extract in PH 9.

Antioxidant status:

At the end of the experiment significant increase ($p < 0.05$) in serum TAO concentration was observed in BCSenPs (T2)

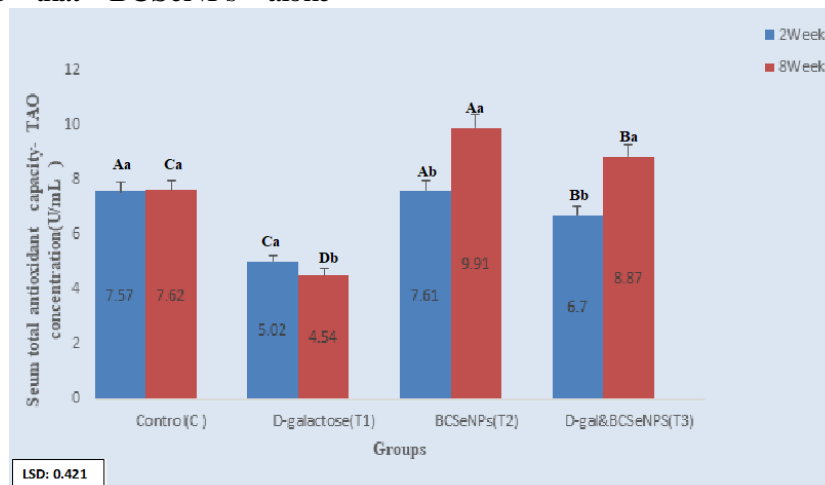
group comparing to the values in D-galactose (T1), D-gal& BCSenPs (T3) and control (C) group (Histogram1). Within the time, significant ($P > 0.05$) differences (increase or

decrease) where observation in all treated groups when compared to each other. Serum malondialdehyde (MDA) concentration (nmol/ml) showed significance elevation ($p < 0.05$) in D-gal(T1) group comparing to the values in the control(C), T2 and T3 groups at the end of experiment. The result also showed that intubated black currant- selenium nanoparticles (BCSeNPs) 1mg/kg B.W. for 8 weeks caused significant decrease ($p < 0.05$) in serum MDA concentration comparing to value in D-gal group (Histogram2). Within the time, D-galactose (T1) and D-gal & BCSeNPs (T3) groups were significant elevation in this parameter when compared to T2 and control groups. Results in histogram (3) indicated a significant ($p < 0.05$) elevation in the fold change of GSH-Px gene expression levels in T2 and T3 groups when compared with T1 and control groups. There was non-significant decreasing the fold change in T 1 group (0.88 ± 0.24) as compared with control group.

Lipid profile:

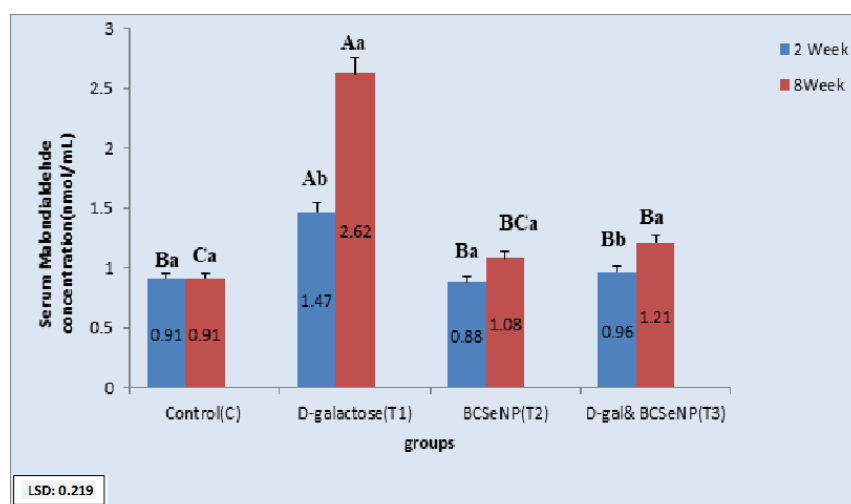
Comparing to control, T2 and T3 treated groups, at the end of the experiment, significant elevation ($p < 0.05$) in serum (TC) was observed in T1 treated group that received 150 mg/kg B.W of D-galactose. The result also showed that BCSeNPs alone

(groupT2), or their combination with D-galactose (groupT3), caused significant decrease in this parameter at week 2&8 comparing to the T1 treated group (Histogram4). Within the time, D-galactose(T1), BCSeNPs (T2) and D-gal & BCSeNPs(T3) groups showed significant elevation in this parameter at 8th comparing to 2nd weeks. Histogram (5) clarified the effect of BCSeNPs, D-galactose or their combination on serum TAG concentration in adult male rats. After 2 weeks, a significant ($P < 0.05$) elevation in this parameter was observed in groups (T1) after *i.p.* of D-galactose (150 mg/kg B.W.) and T3 (combination of D-gal & BCSeNPs) comparing to the value in groups BCSeNPs and control (T2 and C). Further significant elevation ($P < 0.05$) in serum TAG concentration was observed after eight weeks(T1) treated group comparing to values in other treated groups. In the same treated period, intubated black currant- selenium nanoparticles 1 mg/kg B.W. or T3 treated group caused significant decrease ($P < 0.05$) in serum TAG concentration comparing to the value in T1 treated group.



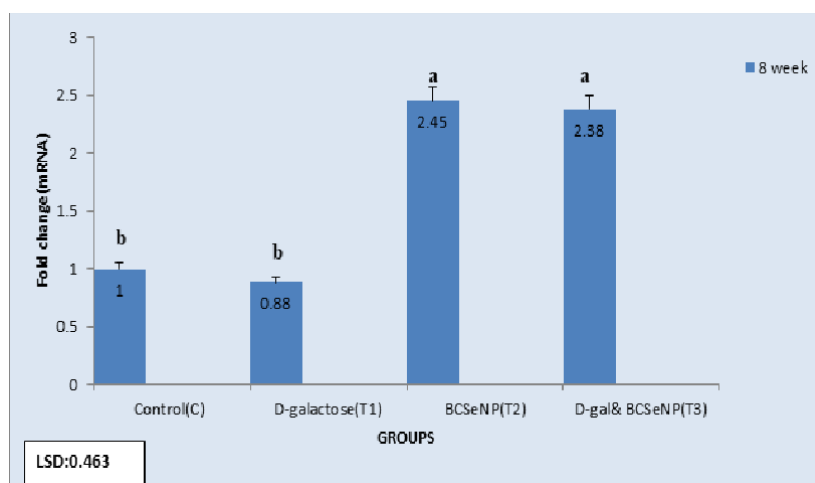
Histogram (1): Effect of D-galactose, Black currant selenium Nanoparticles (BCSeNPs) or their combination eight weeks on serum total antioxidant capacity-TAO concentration (U/mL) in adult rats.

Values are expressed as mean \pm SE. n = 8. Various capital letters denote significant differences ($P < 0.05$) between groups. Various small letters denote significant differences ($P < 0.05$) between periods. Control (C): Intact rats where given daily normal saline (orally and *i.p.*). D-gal (T1): animals in this group were subjected to *i.p.* injection of D gal at a dose (150 mg/kg/day). BCSeNP (T2): rats were intubated black currant- selenium nanoparticles (1mg/Kg.B.W). D-gal & BCSeNP (T3): animals in this group were administered BCSeNPs concurrently with *i.p.* injected of D-gal.



Histogram (2): Effect of D-galactose, Black currant selenium Nanoparticles (BCSeNPs) or their combination eight weeks on serum malondialdehyde concentration (nmol/ml) in adult rats.

Values are expressed as mean \pm SE. n = 8. Various capital letters denote significant differences ($P < 0.05$) between groups. Various small letters denote significant differences ($P < 0.05$) between periods. Control (C): Intact rats where given daily normal saline (orally and *i.p.*). D-gal (T1): animals in this group were subjected to *i.p.* injection of D gal at a dose (150 mg/kg/day). BCSeNP (T2): rats were intubated black currant- selenium nanoparticles (1mg/Kg.B.W). D-gal & BCSeNP (T3): animals in this group were administered BCSeNPs concurrently with *i.p.* injected of D-gal.



Histogram (3): Effect of D-galactose, Black currant selenium Nanoparticles (BCSeNPs) or their combination eight weeks on gene expression of glutathione peroxidase in adult rats.

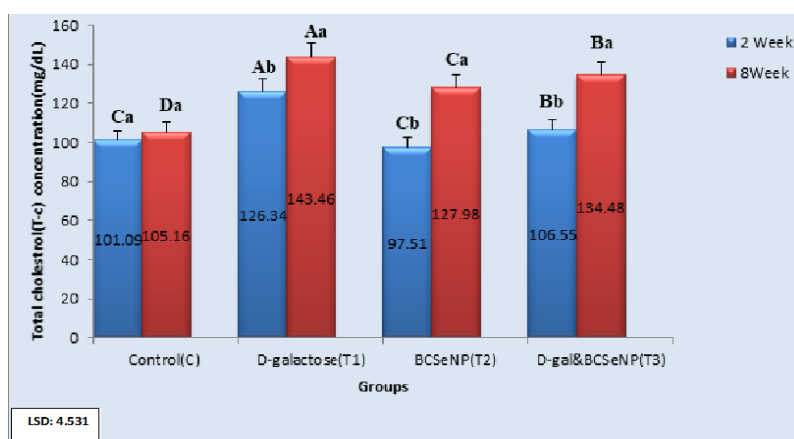
Various small letters denote significant differences ($P < 0.05$) between periods. Control (C): Intact rats where given daily normal saline (orally and *i.p.*). D-galactose (T1): animals in this group were subjected to *i.p.* injection of D gal at a dose (150 mg/kg/day). BCSeNP (T2): rats were intubated black currant- selenium nanoparticles (1mg/Kg.B.W). D-gal & BCSeNP (T3): animals in this group were administered BCSeNPs concurrently with IP injected of D-gal.

Besides, combination of D-gal & BCSeNPs caused significant ($P < 0.05$) decrease in this parameter the value tends to normalize that of the control at the end of the experimental. At the end of the experiment, significant ($p < 0.05$) elevation in serum HDL-c concentration was observed after intubated black currant- selenium nanoparticles (T2) or D-gal & BCSeNPs in group (T₃) comparing to the HDL-c value in D-gal treated group and control group (Histogram 6). Within the time,

significant ($p < 0.05$) increase in this parameter was observed in T1, T2 and T3 groups after eight weeks comparing to the value in two weeks. After two weeks, a significant ($p < 0.05$) decrease in this parameter was observed in groups T2 under the effect of BCSeNPs and T3 groups after *i.p.* of D-galactose (150 mg/kg B.W.) & intubation BCSeNPs comparing to the V-LDL-c value in T1 treated groups which showed significant elevation. Further, significant elevation

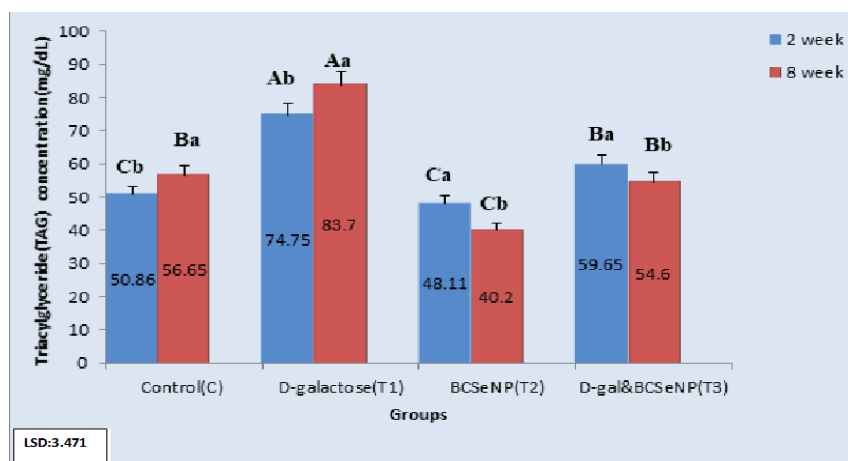
($p < 0.05$) in serum V-LDL-c concentration was observed after eight weeks in D-galactose treated group (T1) comparing to values in other treated groups (Histogram 7). In the same treated period, intubated black currant-selenium nanoparticles (1mg/Kg.B.W) in T2 and T3 caused significant decrease ($p < 0.05$) in serum V-LDL-c concentration comparing to the value in D- gal treated group. The result showed that intubated black currant- selenium nanoparticles (T2) groups caused significant

decrease ($p < 0.05$) in serum LDL-c concentration comparing to the value in other treated groups and the value becomes below that of the control at the end of the experimental. In comparison between periods, control and T3 groups showed no significant ($P > 0.05$) difference between 2nd and 8th weeks periods, whereas T1 and T2 groups recorded significant ($P < 0.05$) increase in the end of experiment (Histogram 8).



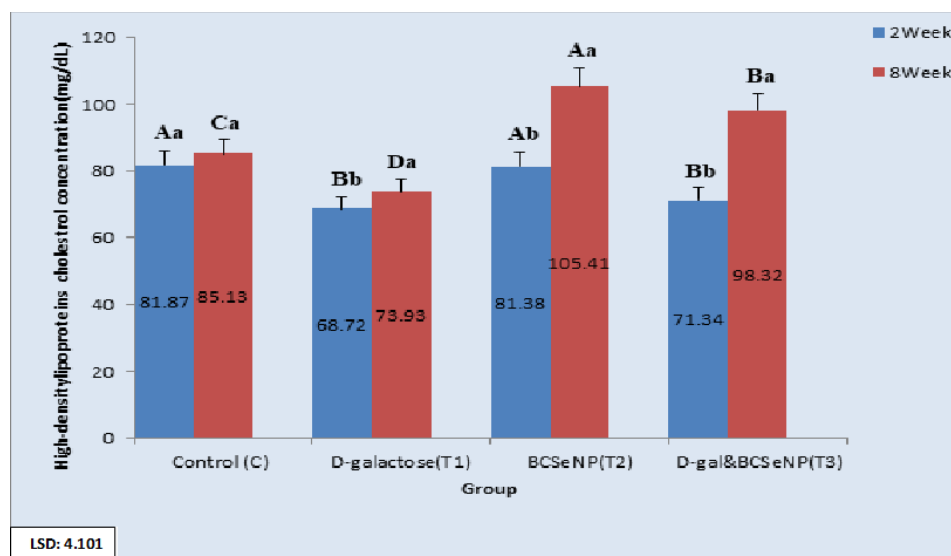
Histogram (4): Effect of D-galactose, Black currant selenium Nanoparticles (BCSeNPs) or their combination eight weeks on serum Total cholesterol concentration (mg/dl) in adult rats.

Values are expressed as mean \pm SE. $n = 8$. Various capital letters denote significant differences ($P < 0.05$) between groups. Various small letters denote significant differences ($P < 0.05$) between periods. Control (C): Intact rats where given daily normal saline (orally and *i.p.*). D-gal (T1): animals in this group were subjected to *i.p.* injection of D gal at a dose (150 mg/kg/day). BCSeNP (T2): rats were intubated black currant- selenium nanoparticles (1mg/Kg.B.W). D-gal & BCSeNP (T3): animals in this group were administered BCSeNPs concurrently with *i.p.* injected of D-gal.



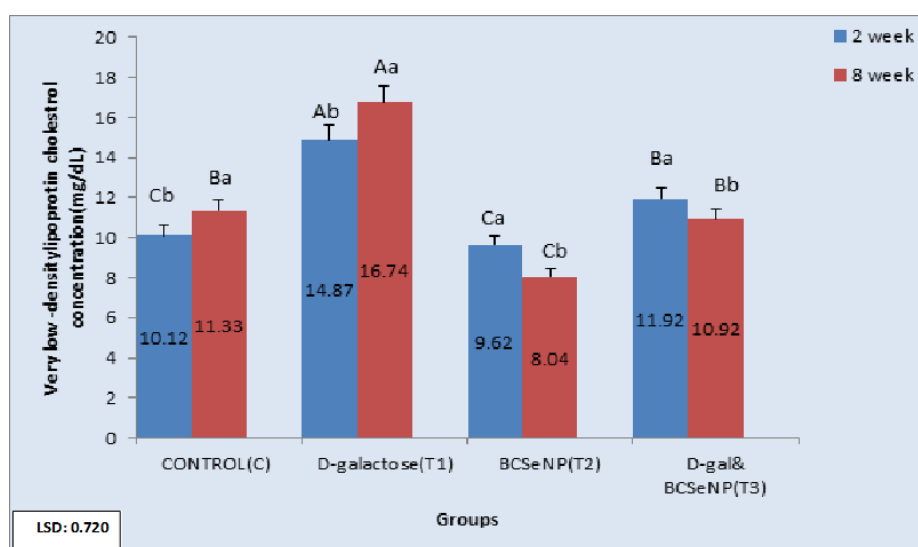
Histogram (5): Effect of D-galactose, Black currant selenium Nanoparticles (BCSeNPs) or their combination eight weeks on triglyceride (TAG) concentration (mg/dl) in adult rats.

Values are expressed as mean \pm SE. $n = 8$. Various capital letters denote significant differences ($P < 0.05$) between groups. Various small letters denote significant differences ($P < 0.05$) between periods. Control (C): Intact rats where given daily normal saline (orally and *i.p.*). D-gal (T1): animals in this group were subjected to *i.p.* injection of D gal at a dose (150 mg/kg/day). BCSeNP (T2): rats were intubated black currant- selenium nanoparticles (1mg/Kg.B.W). D-gal & BCSeNP (T3): animals in this group were administered BCSeNPs concurrently with *i.p.* injected of D-gal.



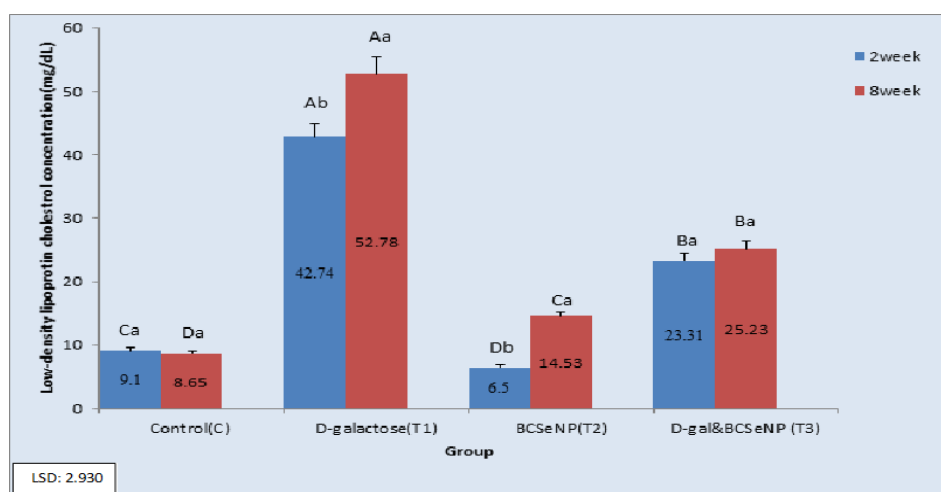
Histogram (6): Effect of D-galactose, Black currant selenium Nanoparticles (BCSeNPs) or their combination eight weeks on serum high-density lipoprotein cholesterol concentration (mg/dL) in adult rats.

Values are expressed as mean \pm SE. n= 8. Various capital letters denote significant differences ($P < 0.05$) between groups. Various small letters denote significant differences ($P < 0.05$) between periods. Control (C): Intact rats where given daily normal saline (orally and *i.p.*). D-gal (T1): animals in this group were subjected to *i.p.* injection of D gal at a dose (150 mg/kg/day). BCSeNP (T2): rats were intubated black currant- selenium nanoparticles (1mg/Kg.B.W). D-gal& BCSeNP (T3): animals in this group were administered BCSeNPs concurrently with *i.p.* injected of D-gal.



Histogram (7): Effect of D-galactose, Black currant selenium Nanoparticles (BcSeNPs) or their combination eight weeks on serum very low-density lipoprotein cholesterol concentration (mg/dl) in adult rats.

Values are expressed as mean \pm SE. n= 8. Various capital letters denote significant differences ($P < 0.05$) between groups. Various small letters denote significant differences ($P < 0.05$) between periods. Control (C): Intact rats where given daily normal saline (orally and *i.p.*). D-gal (T1): animals in this group were subjected to *i.p.* injection of D gal at a dose (150 mg/kg/day). BCSeNP (T2): rats were intubated black currant- selenium nanoparticles (1mg/Kg.B.W). D-gal& BCSeNP (T3): animals in this group were administered BCSeNPs concurrently with *i.p.* injected of D-gal.



Histogram (8): Effect of D-galactose, Black currant selenium Nanoparticles (BCSeNPs) or their combination eight weeks on serum low-density lipoprotein cholesterol concentration (mg/dL) in adult rats.

Values are expressed as mean \pm SE. $n = 8$. Various capital letters denote significant differences ($P < 0.05$) between groups. Various small letters denote significant differences ($P < 0.05$) between periods. Control (C): Intact rats where given daily normal saline (orally and *i.p.*). D-gal (T1): animals in this group were subjected to *i.p.* injection of D gal at a dose (150 mg/kg/day). BCSeNP (T2): rats were intubated black currant- selenium nanoparticles (1mg/Kg.B.W). D-gal& BCSeNP (T3): animals in this group were administered BCSeNPs concurrently with *i.p.* injected of D-gal.

4-Discussion:

Antioxidant status

In current study significant elevation in serum TAO-C concentration, upregulation of GSH-Px gene expression with depression in MDA concentration was observed in BCSeNPs and T3 groups as compared to control and T1 treated groups, which refers to antioxidant effect of SeNPs (9,40). The positive effect of SeNPs could be related to the incorporation of selenium into proteins, such as selenocysteine (SeCys) and its preventive role in oxidative tissue damage (19,41,42). The best way to scavenge ROS by SeNPs could be through the ability to remove potential damage of lipid hydroperoxides and H_2O_2 via upregulation the levels of GSH-Px, SOD, and maintained GSH and productive mitochondrial function (43,44), indicating the antioxidant effect of SeNPs (45,46). Besides, the activity of superoxide dismutase, glutathione peroxidase and glutathione reductase in the serum and liver increased with selenium food supplementation (47). It has been observed that a mixture of selenium nanoparticles-grape seed extract (SeNPs - GSE) possesses antioxidant and anti-diabetic activities by decreasing oxidative stress and scavenger free radicals (9,40). Malondialdehyde is a measure of lipid peroxidation in the tissues, which considered

as one of the important markers of oxidative stress that affect different organs (48). Its elevation by D-galactose in current study indicated oxidative damage induced by D-galactose. Several studies suggest a strong correlation between mitochondrial damage and ROS (mainly H_2O_2) production in cells (49,50). Such elevation in ROS could be accompanied with elevated MDA concentration and intense depletion in TAC (51), as recorded here in D-galactose treated group. Although D-galactose can be changed into glucose at normal concentrations. At high levels, D-galactose is oxidized into aldehydes and hydrogen peroxide, resulting in the generation of ROS and thus LPO product (52) including MDA. In agreement with our results, others observed that subcutaneous injection of D-galactose caused a significant decrease in antioxidant enzyme activities of CAT, GSH-Px, SOD, and T-AOC, as well as an increase in the MDA level (53,54).

lipid profile

Black currant selenium nanoparticles in here in study has been found to be inversely correlated to a case of dyslipidemia and positively correlated to HDL- cholesterol concentration. Similar to our results, in rats Se supplementation has been reported to increase low-density lipoprotein (LDL) receptor activity (55), lowered serum total cholesterol

(56). Alleviation of hyperlipidemic by SeNPs or selenium has been documented (57-59). On the contrary, Hunge and his worker (60), recorded that SeNPs supplementation is independently associated with dyslipidemia. The hypolipidemia induced by SeNPs could be through their efficacy in lowering gene expression of many enzymes associated with hepatic cholesterol metabolic especially HMG-COA reductase, cholesterol storage as well enhancing conversion of cholesterol to bile acids (55, 59,61). It is worth to mention that apart from its lipid lowering activity, selenium could also alleviate hyperlipidemia by reducing oxidative stress, through antioxidant seleno proteins enzyme especially GSH-peroxidase which play an important role in lipid metabolism (59,61). Similarly, present works showed a decrease in oxidative stress depression in MDA and elevation in TOA level and GSH-Px gene expression. Glucose and lipid metabolism disorder have been considered as important factor to D-galactose induced aging (62). Besides, D-galactose induced cardiac hypertrophy (53) has been reported. A substantial amount of evidence has demonstrated that ROS and AGEs, produced after high concentration of D-gal have been implicated in the pathological processes of age-related disease such as diabetes, arteriosclerosis (63,64) in which high lipid profile is regarded as major risk factor. Bo-Htay and his coworker (65) indicated that high concentration of D-gal aggravated cardiac disfunction and hyperlipidemia in rat with high fat diet. An elevation in mitochondrial ROS due to excess galactose caused generation of super oxide anion, decreased membrane potential, decrease lipid metabolism leading to dyslipidemia (66-68), where impairment of mitochondrial function, oxidative metabolism and depression in ATP generation (69,70) could occurred. Besides, oxidative stress induced by excess mitochondrial ROS, caused inhibition of insulin signaling and development of insulin resistance (71) that caused disturbance in lipid metabolism and dyslipidemia (72,73).

Reference:

- 1- Prasad, M.; Lambe, U. P.; Brar, B.; Shah, I.; Manimegalai, J.; Ranjan, K.; Rao, R.; Kumar, S.; Mahant, S.; Khurana, S.K.; Iqbal, H.M.N.; Dhama, K.; Misri, J. and Prasad, G. (2018). Nanotherapeutics: an insight into healthcare and multi-dimensional applications in medical sector of the modern world. *Biomedicine & Pharmacotherapy*, 97: 1521-1537.
- 2- Ebrahimi, K.; Shiravand, S. and Mahmoudvand, H. (2017). Biosynthesis of copper nanoparticles using aqueous extract of Capparis spinosa fruit and investigation of its antibacterial activity. *Marmara Pharmaceutical Journal* 21/4: 866- 871.
- 3- Khatami, M.; Amini, E.; Amini, A.; Mortazavi, S.M.; Kishani Farahani, Z. and Heli, H. (2017). Biosynthesis of silver nanoparticles using pine pollen and evaluation of the antifungal efficiency. *Iran J Biotechnol.* 15: 1–7.
- 4- Husen, A. and Siddiqi, K. S. (2014). Plants and microbes assisted selenium nanoparticles: characterization and application. *Journal of Nanobiotechnology*, 12:1–10.
- 5- Husain, W.M.; Araak, J.K. and Ibrahim, O.M. (2019). Green Synthesis of Zinc Oxide Nanoparticles from (Punica granatum L) Pomegranate Aqueous Peel Extract. *Iraqi Journal of Veterinary Medicine (Iraqi J. Vet. Med.)* p-ISSN:1609-5693, e-ISSN:2410-7409, 43(2), 6-14. <https://doi.org/10.30539/iraqijvm.v43i2.524>.
- 6- Zhang, J.; Wang, X. and Xu, T. (2008). Elemental selenium at nano size (Nano-Se) as a potential chemopreventive agent with reduced risk of selenium toxicity: comparison with Se-methylselenocysteine in mice. *Toxicol Sci.* 101(1):22–31.
- 7- Fairweather-Tait, S.J.; Bao, Y.; Broadley, M.R.; Collings, R.; Ford, D.; Hesketh, J.E. and Hurst, R. (2011). Selenium in human health and disease. *Antioxid Redox Signal*, 14: 1337-1383.
- 8- Krishnan, V.; Loganathan, C. and Thayumanavan, P. (2019). Green synthesized selenium nanoparticles using

- Spermatozoa of *hispidus* as carrier of s-allyl glutathione: to accomplish hepatoprotective and nephroprotective activity against acetaminophen toxicity. *Artificial cells, nanomedicine, and biotechnology*, 47(1): 56-63.
- 9- **Abdelaleem, R. M.A.; Abdel Hameed, H.F.; Askar, M.E.; Hassan, S.H.M. and El-Bata, A.I. (2016).** Modulatory Role of Selenium Nanoparticles and Grape Seed Extract Mixture on Oxidative Stress Biomarkers in Diabetic Irradiated Rats. *Indian Journal of Pharmaceutical Education and Research*, 50 (1):170-178.
- 10- **Sakr, T. M.; Korany, M. and Katti, K. V. (2018).** Selenium nanomaterials in biomedicine—An overview of new opportunities in nanomedicine of selenium. *Journal of Drug Delivery Science and Technology*, 46: 223-233.
- 11- **Khurana, A.; Tekula, S.; Saifi, M. A.; Venkatesh, P. and Godugu, C. (2019).** Therapeutic applications of selenium nanoparticles. *Biomedicine & Pharmacotherapy*, 111: 802–812.
- 12- **Djanaguiraman, M.; Belliraj, N.; Bossmann, S. and Prasad, P. (2018).** High-Temperature Stress Alleviation by Selenium Nanoparticle Treatment in Grain Sorghum. *ACS Omega*, 3(3): 2479–2491.
- 13- **Yu, Y.; Bai, F.; Wang, W.; Liu, Y.; Yuan, Q.; Qu, S.; Zhang, T.; Tian, G.; Li, S.; Li, D. and Ren, G. (2015).** Fibroblast growth factor 21 protects mouse brain against D-galactose induced aging via suppression of oxidative stress response and advanced glycation end products formation. *Pharmacology biochemistry and behavior*, 133: 122-131.
- 14- **Li, L.; Xu, M.; Shen, B.; Li, M.; Gao, Q. and Wei, S. G. (2016).** Moderate exercise prevents neurodegeneration in D-galactose-induced aging mice. *Neural Regeneration Research*, 11:807–815.
- 15- **Munch, G.; Westcott, B.; Menini, T.; Gugliucci, A. (2012).** Advanced glycation endproducts and their pathogenic roles in neurological disorders. *Amino Acids*, 42:1221–1236.
- 16- **Zhang, X.; Jin, C.; Li, Y.; Guan, S.; Han, F. and Zhang, S. (2013).** Catalpol improves cholinergic function and reduces inflammatory cytokines in the senescent mice induced by D-galactose. *Food and Chemical Toxicology*, 58: 50-55.
- 17- **Mallidis, C.; Agbaje, I.; Rogers, D.; Glenn, J.; McCullough, S.; Atkinson, A.B.; Steger, K.; Stitt, A. and McClure, N. (2007).** Distribution of the receptor for advanced glycation end products in the human male reproductive tract: prevalence in men with diabetes mellitus. *Human Reproduction*, 22(8):2169-2177.
- 18- **Ali, T.; Badshah, H.; Kim, T. H., and Kim, M. O. (2015).** Melatonin attenuates D-galactose-induced memory impairment, neuroinflammation and neurodegeneration via RAGE/NF-KB/JNK signaling pathway in aging mouse model. *Journal of pineal research*, 58(1): 71-85.
- 19- **Bai, K.; Hong, B.; Hong, Z.; Sun, J. and Wang, C. (2017).** Selenium nanoparticles-loaded chitosan/citrate complex and its protection against oxidative stress in D-galactose-induced aging mice. *Journal of nanobiotechnology*, 15(1): 92.
- 20- **Jeremy, M.; Gurusubramanian, G. and Roy, V. K. (2017).** Localization pattern of visfatin (NAMPT) in d-galactose induced aged rat testis. *Annals of Anatomy-Anatomischer Anzeiger*, 211: 46-54.
- 21- **Sulistyoningrum, E. (2017).** D-galactose-induced animal model of male reproductive aging. *Jurnal Kedokteran dan Kesehatan Indonesia*, 8(1): 19-28.
- 22- **Li, M.; Wang, S.; Li, X.; Jiang, L.; Wang, X.; Kou, R.; Wang, Q.; Xu, L.; Zhao, N. and Xie, K. (2018).** Diallyl sulfide protects against lipopolysaccharide/D-galactosamine-induced acute liver injury by inhibiting oxidative stress, inflammation and apoptosis in mice. *Food and Chemical Toxicology*, 120: 500–509.
- 23- **Saleh, D.O.; Mansour, D.F.; Hashad, I.M. and Bakeer, R.M. (2019).** Effects of sulfuraphane on D-galactose-induced liver aging in rats: role of keap-1/nrf-2 pathway. *Eur J Pharmacol*, 855:40–49.

- 24- **Gottimukkala, K.; Harika, R. and Deeveka, Z. (2017)**. Green synthesis of iron nanoparticles using green tea leaves extract. *Nanomedicine Biotherapeutic Discovery*, 7: 1 – 5.
- 25- **Kaikai, B.; Bihong, H.; Jianlin, H.; Zhuan, H. and Ran, T. (2017)**. Preparation and antioxidant properties of selenium nanoparticles-loaded chitosan microspheres *International Journal of Nanomedicine*, 12: 4527–4539.
- 26- **Mittal, A. K.; Chaisti, Y. and Banerjee, U. C. (2013)**. Synthesis of metallic nanoparticles using plant extracts. *Biotechnol. Adv.*, 31:346-356.
- 27- **Banerjee, P.; Satapathy, M.; Mukhopahayay, A. and Das, P. (2014)**. leaf extract mediated green synthesis of silver nanoparticles from widely available indian plants: synthesis, characterization, antimicrobial property and toxicity analysis. *Bioresour. Bioprocess*, 1: 3.
- 28- **Rietveld, H. (1969)**. A profile refinement method for nuclear and magnetic structures. *J. Appl. Crystallogr.*, 2:65-71.
- 29- **Holzwarth, U. and Gibson, N. (2011)**. The scherrer equation versus the Debye-Scherrer equation. *Nat. Nanotechnol.*, 6:534.
- 30- **Khoshnamvand, M.; Huo, C. and Liu, J. (2018)**. Silver nanoparticles synthesis using *Allium ampeloprasum* L. leaf extract: characterization and performance in catalytic reduction of 4-nitrophenol and antioxidant activity. *J. Mol. Struct.*, 1175:90-96.
- 31- **Mittal, A. K.; Tripathy, D.; Choudhary, A.; Aili, P. K.; Chatterjee, A.; Singh, I. P. and Banerjee, U. C. (2015)**. Bio-synthesis of silver nanoparticles using *Potentilla fulgens* Wall. ex Hook. and its therapeutic evaluation as anticancer and antimicrobial agent. *Materials Science and Engineering: C*, 53: 120-127.
- 32- **Rolim, W. R.; Pelegrino, M. T.; de Araújo Lima, B.; Ferraz, L. S.; Costa, F. N.; Bernardes, J. S.; Rodrigues, T.; Brocchi, M. and Seabra, A. B. (2019)**. Green tea extract mediated biogenic synthesis of silver nanoparticles: Characterization, cytotoxicity evaluation and antibacterial activity. *Applied Surface Science*, 463: 66-74.
- 33- **Allain, C. C.; Poon, L. S.; Chan, C. S.; Richmond, W. F. P. C. and Fu, P. C. (1974)**. Enzymatic determination of total serum cholesterol. *Clinical chemistry*, 20(4): 470-475.
- 34- **Kaplan, A. and Lee, V. F. (1965)**. Serum lipid levels in infants and mothers at parturition. *Clinica chimica acta*, 12(3): 258-263.
- 35- **Friedewald, W. T.; Levy, R. I. and Fredrickson, D. S. (1972)**. Estimation of the concentration of low-density lipoprotein cholesterol in plasma, without use of the preparative ultracentrifuge. *Clinical chemistry*, 18(6): 499-502.
- 36- **Naito, H. K. and Kaplan, A. (1984)**. High-density lipoprotein (HDL) cholesterol. *Clin Chem. Toronto. Princeton*, 1207-13.
- 37- **Buege, J. A. and Aust, S. D. (1978)**. Microsomal lipid peroxidation. In *Methods in enzymology*, 52:302-310.
- 38- **Livak, K. J. and Schmittgen, T. D. (2001)**. Analysis of relative gene expression data using real-time quantitative PCR and the 2- $\Delta\Delta CT$ method. *methods*, 25(4): 402-408.
- 39- **Snedecor George, W. and Cochran, W. G. (1973)**. *Statistical methods*. Iowa State University Press.
- 40- **Al-Othman, A.M.; Al-Numair, S.; El-Desoky, K.G.; Yusuf, K.; Al Othman, Z.A.; Aboul-Soud, M.A. and Giesy, J. P. (2011)**. Protection of α -tocopherol and selenium against acute effects of Malathion on liver and kidney of rats. *Afr. J. Pharm. Pharmacol*, 5(10): 1263-1271.
- 41- **Kojouri, G. A. and Sharifi, S. (2013)**. Preventing effects of nano-selenium particles on serum concentration of blood urea nitrogen, creatinine, and total protein during intense exercise in donkey. *Journal of equine veterinary science*, 33(8): 597-600.
- 42- **Zheng, S.; Song, H.; Gao, H.; Liu, C.; Zhang, Z. and Fu, J. (2016)**. The antagonistic effect of selenium on lead-induced inflammatory factors and heat

- shock protein mRNA level in chicken cartilage tissue. *Biological trace element research*, 173(1): 177-184.
- 43- **Rowntree, J.E.; Hill, G.M. and Hawkins, D.R. (2004)**. Effect of Se on selenoprotein activity and thyroid hormone metabolism in beef and dairy cows and calves. *J Anim Sci.*,82:2995–3005.
- 44- **Bai, K.; Hong, B.; Huang, W. and He, J. (2020)**. Selenium-Nanoparticles-Loaded Chitosan/Chitooligosaccharide Microparticles and Their Antioxidant Potential: A Chemical and In Vivo Investigation. *Pharmaceutics*, 12(1): 43.
- 45- **Dkhil, M. A.; Zrieq, R.; Al-Quraishy, S. and Abdel Moneim, A. E. (2016)**. Selenium nanoparticles attenuate oxidative stress and testicular damage in streptozotocin-induced diabetic rats. *Molecules*, 21(11):1517.
- 46- **Zhu, K.; Zeng, X.; Tan, F.; Li, W.; Li, C.; Song, Y. and Zhao, X. (2019)**. Effect of insect tea on D-galactose-induced oxidation in mice and its mechanisms. *Food Science & Nutrition*, 7(12): 4105-4115.
- 47- **Wang, L.; Xiao, J.; Hua, Y.; Xiang, X.; Zhou, Y.; Ye, L. and Shao, Q. (2019)**. Effects of dietary selenium polysaccharide on growth performance, oxidative stress and tissue selenium accumulation of juvenile black sea bream, *Acanthopagrus schlegelii*. *Aquaculture*, 503: 389–395.
- 48- **Hassan, Q.; Akhtar, M.; Ahmed, S.; Ahmad, A. and Najmi, A.K. (2017)**. *Nigella sativa* protects against isoproterenol-induced myocardial infarction by alleviating oxidative stress, biochemical alterations and histological damage. *Asian Pac. J. Trop. Biomed.*, 7(4): 294–299.
- 49- **Chairuangkitti, P.; Lawanprasert, S.; Roytrakul, S.; Aueviriyavit, S.; Phummiratch, D.; Kulthong, K.; Chanvorachote, P. and Maniratanachote, R. (2013)**. Silver nanoparticles induce toxicity in A549 cells via ROS-dependent and ROS-independent pathways. *Toxicol. Vitr.*, 27(1): 330–338.
- 50- **Christiansen, L.B.; Dela, F.; Koch, J.; Hansen, C.N.; Leifsson, P.S. and Yokota, T. (2015)**. Impaired cardiac mitochondrial oxidative phosphorylation and enhanced mitochondrial oxidative stress in feline hypertrophic cardiomyopathy. *Am. J. Physiol. Circ. Physiol.*, 308(10): H1237–H1247
- 51- **Sies, H. (2017)**. Hydrogen peroxide as a central redox signaling molecule in physiological oxidative stress: Oxidative eustress. *Redox biology*, 11: 613-619.
- 52- **Mohammadi, E.; Mehri, S.; Bostan, H. B. and Hosseinzadeh, H. (2018)**. Protective effect of crocin against d-galactose-induced aging in mice. *Avicenna journal of phytomedicine*, 8(1): 14.
- 53- **Chang, Y.M.; Chang, H.H.; Lin, H.J.; PadmaViswanadha, V.; Chen, R.J. and Huang, C.Y. (2017)**. Inhibition of cardiac hypertrophy effects in D-galactose-induced senescent hearts by alpinate oxyphyllae fructus treatment. *Evid Based Complement Alternat Med.*,2017: 2624384.
- 54- **Liang, C. Y.; Liang, Y. M.; Liu, H. Z.; Zhu, D. M.; Hou, S. Z.; Wu, Y. Y.; Huang, S. and Lai, X. P. (2017)**. Effect of *Dendrobium officinale* on D-galactose-induced aging mice. *Chinese journal of integrative medicine*,12:1-9.
- 55- **Dhingra, S. and Bansal, M. (2006)**. Attenuation of LDL receptor gene expression by selenium deficiency during hypercholesterolemia. *Mol Cell Biochem*, 282: 75–82.
- 56- **Bunglavan, S. J.; Garg, A. K.; Dass, R. S. and Shrivastava, S. (2014)**. Effect of supplementation of different levels of selenium as nanoparticles/sodium selenite on blood biochemical profile and humoral immunity in male Wistar rats. *Veterinary World*, 7(12):1075-1081.
- 57- **Hasani, M.; Djalalinia, S.; Sharifi, F.; Varmaghani, M.; Zarei, M.; Abdar, M. E.; Asayesh, H.; Noroozi, M.; Kasaeian, A.; Gorabi, A.M. and Qorbani, M. (2018)**. Effect of selenium supplementation on lipid profile: a systematic review and meta-analysis.

- Hormone and Metabolic Research, 50(10):715-727.
- 58- **Abdulmalek, S. A. and Balbaa, M. (2019)**. Synergistic effect of nano-selenium and metformin on type 2 diabetic rat model: Diabetic complications alleviation through insulin sensitivity, oxidative mediators and inflammatory markers. *PloS one*, 14(8).
- 59- **Guo, B.; Guo, Q.; Wang, Z.; Shao, J. B.; Liu, K.; Du, Z. D. and Gong, S. S. (2020)**. D-Galactose-induced oxidative stress and mitochondrial dysfunction in the cochlear basilar membrane: an in vitro aging model. *Biogerontology*, 1-13.
- 60- **Huang, Y. Q.; Shen, G.; Lo, K., Huang, J. Y.; Liu, L.; Chen, C. L.; Yu, Y.L.; Sun, S.; Zhang, B. and Feng, Y. Q. (2020)**. Association of circulating selenium concentration with dyslipidemia: Results from the NHANES. *Journal of Trace Elements in Medicine and Biology*, 58, 126438.
- 61- **Hamza, R. Z.; EL-Megharbel, S. M.; Altalhi, T., Gobouri, A. A. and Alrogi, A. A. (2020)**. Hypolipidemic and hepatoprotective synergistic effects of selenium nanoparticles and vitamin. E against acrylamide-induced hepatic alterations in male albino mice. *Applied Organometallic Chemistry*, e5458.
- 62- **Ahangarpour, A.; Oroojan, A.A. and Heidari, H. (2014)**. Effects of Exendin-4 on LH, FSH, testosterone and sperm count of aged mice model induced by D-galactose. *World J Mens Health*. 32(3):176-183.
- 63- **Byun, K.; Yoo, Y.; Son, M.; Lee, J.; Jeong, G. B.; Park, Y. M.; Salekdeh, G.H. and Lee, B. (2017)**. Advanced glycation end-products produced systemically and by macrophages: A common contributor to inflammation and degenerative diseases. *Pharmacology & therapeutics*, 177: 44-55.
- 64- **Fitri, S.; Anggraini, D. R. and Ichwan, M. (2020)**. Effects of Gambir leaves extract (*Uncaria gambir* Roxb.) in preventing the aging process induced D-galactose on pancreas mice. In *IOP Conference Series: Earth and Environmental Science*, 425(1): 012021.
- 65- **Bo-Htay, C., Shwe, T., Higgins, L., Palee, S., Shinlapawittayatorn, K., Chattipakorn, S. C., & Chattipakorn, N. (2020)**. Aging induced by D-galactose aggravates cardiac dysfunction via exacerbating mitochondrial dysfunction in obese insulin-resistant rats. *GeroScience*, 42(1): 233-249.
- 66- **Houstis, N.; Rosen, E. D. and Lander, E. S. (2006)**. Reactive oxygen species have a causal role in multiple forms of insulin resistance. *Nature*, 440(7086):944-948.
- 67- **Ohta, Y.; Kinugawa, S.; Matsushima, S.; Ono, T.; Sobirin, M. A.; Inoue, N.; Yokota, T.; Hirabayashi, K. and Tsutsui, H. (2011)**. Oxidative stress impairs insulin signal in skeletal muscle and causes insulin resistance in postinfarct heart failure. *American Journal of Physiology-Heart and Circulatory Physiology*, 300(5): H1637-H1644.
- 68- **Xu, F.; Liu, Y.; Zhao, H.; Yu, K.; Song, M.; Zhu, Y. and Li, Y. (2017)**. Aluminum chloride caused liver dysfunction and mitochondrial energy metabolism disorder in rat. *Journal of inorganic biochemistry*:174, 55-62.
- 69- **Nisoli, E.; Clementi, E.; Carruba, M. O. and Moncada, S. (2007)**. Defective mitochondrial biogenesis: a hallmark of the high cardiovascular risk in the metabolic syndrome. *Circulation research*, 100(6): 795-806.
- 70- **Bhatti, J. S.; Bhatti, G. K. and Reddy, P. H. (2017)**. Mitochondrial dysfunction and oxidative stress in metabolic disorders—A step towards mitochondria based therapeutic strategies. *Biochimica et Biophysica Acta (BBA)-Molecular Basis of Disease*, 1863(5): 1066-1077.
- 71- **Kenawy, S.; Hegazy, R.; Hassan, A.; El-Shenawy, S.; Gomaa, N.; Zaki, H. and Attia, A. (2017)**. Involvement of insulin resistance in D-galactose-induced age-related dementia in rats: Protective role of metformin and saxagliptin. *PloS one*: 12(8).

- 72- **Choksi, K. B.; Boylston, W. H.; Rabek, J. P.; Widger, W. R. and Papaconstantinou, J. (2004).** Oxidatively damaged proteins of heart mitochondrial electron transport complexes. *Biochimica et Biophysica Acta (BBA)-Molecular Basis of Disease*, 1688(2): 95-101.
- 73- **Morino, K.; Petersen, K. F.; Dufour, S.; Befroy, D.; Frattini, J.; Shatzkes, N.; Neschen, S.; White, M.F.; Bilz, S.; Sono, S. and Pypaert, M. (2005).** Reduced mitochondrial density and increased IRS-1 serine phosphorylation in muscle of insulin-resistant offspring of type 2 diabetic parents. *The Journal of clinical investigation*, 115(12): 3587-3593